

# CHOCOLATE AGAR WITH BIOVITEX

## Intended Use

**Chocolate Agar** is used with hemoglobin and enrichment for the isolation and cultivation of *Haemophilus influenzae* and other fastidious organisms.

## Principles of the Procedure

Enzymatic Digest of Casein and Enzymatic Digest of Animal Tissue provide nitrogen, carbon, and minerals in GC Agar. Corn Starch absorbs any toxic metabolites produced. The Phosphates are buffering agents. Sodium Chloride maintains osmotic balance of the medium. Agar is the solidifying agent. Chocolate Agar is prepared from GC Agar with the addition of Hemoglobin. Hemoglobin provides hemin (X factor) required for growth of *Haemophilus* and enhanced growth of *Neisseria* spp. A chemical enrichment composed of cofactors, vitamins, and nicotinamide adenine dinucleotide (NAD) are also required for growth of *Haemophilus* and *Neisseria* spp. If required, antimicrobial supplements are added as inhibitors for improved selectivity of the medium.

## Formula / Liter

Enzymatic Digest of Casein.....	7.5 g
Enzymatic Digest of Animal Tissue.....	7.5 g
Corn Starch.....	1.0 g
Dipotassium Phosphate.....	4.0 g
Monopotassium Phosphate.....	1.0 g
Sodium Chloride.....	5.0 g
Agar.....	10.0 g

## Supplements:

Hemoglobin Powder, 10 g

Growth Enrichment, 2 mL

Final pH: 7.2 ± 0.2 at 25°C

## Precautions

1. For Laboratory Use.

## Directions

1. Suspend 36,0 g of the GC Agar in 500 mL of purified water.
2. Heat with frequent agitation and boil for one minute to completely dissolve the medium.
3. Autoclave at 121°C for 15 minutes. Cool to 45 – 50°C.
4. Suspend 10,0 g of hemoglobin powder in 500 mL of purified water and autoclave at 121°C for 15 minutes.
5. Cool to 45 - 50°C and aseptically add to the molten GC Agar. Add 2 mL of growth enrichment. Mix thoroughly and dispense.

**Dehydrated Appearance:** Powder is homogeneous, free flowing, and light beige.

**Prepared Appearance:** Prepared Chocolate Agar is opaque and brown.

**Expected Cultural Response:** Cultural response on Chocolate Agar at 35°C under CO<sub>2</sub> enrichment after 18 – 24 hours incubation.

Microorganism	Response
<i>Haemophilus influenza</i> ATCC® 10211	growth
<i>Neisseria gonorrhoeae</i> ATCC® 43070	growth
<i>Neisseria meningitidis</i> ATCC® 13090	growth
<i>Streptococcus agalactiae</i> ATCC® 13813	growth
<i>Streptococcus pneumoniae</i> ATCC® 6303	growth

## Test Procedure

For a complete discussion on the isolation and identification of *Neisseria* spp. and *Haemophilus* spp. consult procedures outlined in the references.<sup>8,9</sup>

## Results

Refer to appropriate references and procedures for results.

**Storage** ready to use plates - 6-12°C

**Packaging** cat No. 1080 ready to use plates Ø 90 mm (1 x 10 pcs);

**Expiration** ready to use plates - 90 days

### Limitation of the Procedure

Although certain diagnostic tests may be performed directly on Chocolate Agar, biochemical and immunological testing using pure cultures are recommended for complete identification.

### References

1. **Johnson, J.** 1945. Comparison of gonococcus cultures read at 24 and 48 hours. J. Venera. Dis. Inform. **26**:239.
2. **Lankford, C. E., V. Scott, M. F. Cox, and W. R. Cooke.** 1943. Some aspects of nutritional variation of the gonococcus. J. Bacteriol. **45**:321.
3. **Thayer, J. D., and J. E. Martin, Jr.** 1966. Improved medium selective for cultivation of *N. gonorrhoeae* and *N. meningitidis*. Public Health Rep. **81**:559.
4. **Thayer, J. D., and A. Lester.** 1971. Transgrow, a medium for transport and growth of *Neisseria gonorrhoeae* and *Neisseria meningitidis*. HSMHA Health Service Rep. **86**:30.
5. **Martin, J. E., Jr., and R. L. Jackson.** 1975. A biological environmental chamber for the culture of *N. gonorrhoeae* with a new commercial medium. Public Health Rep. **82**:361.
6. **Martin, J. E., Jr., and J. S. Lewis.** 1977. Anisomycin: improved anti-mycotic activity in modified Thayer-Martin Medium. Public Health Rep. **35**:53.
7. **MacFaddin, J. F.** 1985. Media for isolation-cultivation-identification-maintenance of medical bacteria, vol. 1, Williams & Wilkins, Baltimore, MD.
8. **Isenberg, H. D. (ed.).** 1992. Clinical microbiology procedures handbook. vol. 1. American Society for Microbiology, Washington, D.C.
9. **Murray, P. R., E. J. Baron, M. A. Pfaller, F. C. Tenover, and R. H. Tenover (eds.).** 1995. Manual of clinical microbiology, 6<sup>th</sup> ed. American Society for Microbiology, Washington, D.C.